







Test Report

Sample Name:

PLGA

Client Name:

eSUNMed Biotechnology (Shenzhen) Co.,

Ltd.

Client Address:

3F, No. 9, Yifeng Hua Innovation Industrial

Park, Xinshi Community, Dalang Street,

Longhua District, Shenzhen

Test

Items:

MTT cytotoxicity test

Date of Issue:

2025.04.17

Shanghai WEIPU Testing Technology Group Co., LTD.



DECLARE

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Task No.	1		ection egory	Commission test	
Sample No.	BP-S-2501388	5 San	nple rce	Sent by client	
Sample name	PLGA	PI GA5050			
Specification	50:50	Sample quantity 1pc			
Model	1				
Manufacturer	eSUNMed Bio office	technology (Shenz	then) Co.,	Ltd. Wuhan branch	
Manufacturer address		o. 24, Gold-Industria		Hi-tech International ad, Zhengdian Street,	
Client	eSUNMed Biot	echnology (Shenzhe	en) Co., Ltd	.	
Client address		Yifeng Hua Innov lang Street, Longhu		ustrial Park, Xinshi Shenzhen	
Receiving date	2025.03.05				
Test location	3rd Floor, Building 7,166-1, Fengjin Road, Fengxian District, Shanghai.				
Test period	2025.03.05 to 2	2025.03.18			
Test item	MTT cytotoxicit	y test			
Test criterion	ISO 10993-5:20	009			
Test conclusion		y grade is 0, the		is 106.16%, and the ract had no potential	
Implementati on standard	ISO/IEC 17025: 2017; RB/T214—2017				
Remarks		indicates that this it	tem is blank	COST GROUP CO. LID	
Edited by		Checked by	(Au	Approved by the uthorized signatory)	
7	7N	1 0) E EAD	



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1 Objective

The biological response to L-929 cells was evaluated by in vitro cytotoxicity test.

2 Test method

MTT cytotoxicity test

3 Test and control samples

3.1 Test samples

The information in the form is provided by the client.

Sample name	PLGA
Sterilization state	Unsterilized
Sterilization methods	
Sample material	PLGA
Application	1

3.2 Control samples

Negative control sample	e: HDPE		
Manufacturer USP			
Specification	Three-piece pack		
Batch No. R149K0			
Positive control sample:	DMSO		
Manufacturer	Sinopharm Chemical Reagent Co., Ltd.		
Specification	500mL/bottle		
Batch No.	20230922		
Blank control sample:	The MEM medium contained 10% FBS		

4 Reagents and Instrument

4.1 Reagents

Name	Supplier		
FBS	Bio-Channel		
1×MEM medium (100IU/mL PNC, 100µg/mL Streptomycin)	Bio-Channel		
Trypsin (EDTA) solution	Gibco		
PBS	Biosharp		



Name	Supplier		
MTT	Beyotime		
IPA	Sinopharm Chemical Reagent Co., Ltd.		

4.2 Instrument

Name	Instrument ID	Calibration is valid until		
Clean bench	WPE-TL0125	2025.10.09		
Electronic scales	WPE-TL0242	2025.04.10		
Double plate constant temperature oscillation incubator	WPE-TL0390	2025.10.09		
Biological microscope	WPE-TL0139	1		
Centrifuge	WPE-TL0279	2025.04.10		
Microplate reader	WPE-TL0293	2025.10.09		
CO ₂ incubator	WPE-TL0077	2025.04.10		
pH meter	WPE-TL0394	2025.11.20		
Steel ruler	WPE-TL0033	2025.07.18		

5 Test system

Cloning L929 is standard recommended cell line, and this cell comes from Cell Bank/Stem Cell Bank, Chinese Academy of Sciences.

Contact of the test sample with the test system via an extract solution (The MEM medium contained 10% FBS) is considered the optimal route of administration and is the recommended method in standard.

6 Experimental content

6.1 Sterilization

Petri dishes, porous culture plates, pipette tips, and other utensils that may be used in the test are sterilized by high pressure steam prior to the test.

6.2 Sample preparation

Under aseptic operation, the extracts were prepared according to the method in the table below. After the extraction, the changes of the extracts were checked. The extracts were not centrifuged and etc. The pH was not adjusted. Blank control, negative control and positive control samples were prepared by the same method.



Table 6-1 Preparation of extracts

Extraction solvent	Actually sample	Sampling ratio	Solvent volume	Sampling condition	Whether it is clear	рН
MEM medium containing 10%FBS	2.0129g	0.2g:1mL	13.13mL ^a	37℃ 24h 60rpm	Yes	8.00

a: Sample absorption rate is 1.53mL/g.

6.3 Test procedure

The test procedure is sterile operation.

L929 monolayer cells cultured in 10% FBS MEM medium for 48 h to 72 h were liquefied with enzyme liquid (trypsin / EDTA).

The cells are then resuspended in culture medium and the cell suspension is adjusted at a density of 1×10⁵ cells/mL.

Using a multichannel pipette, dispense 100 μ l culture medium only (blank) into the peripheral wells of a 96-well tissue culture microtitre plate. In the remaining wells, dispense 100 μ l of a cell suspension of 1×10⁵ cells/mL. Set blank (left and right 2 groups), negative control, positive control, sample group, each group has 6 parallel wells.

Incubate cells for 24 h (5% CO₂, 37° C, > 90% humidity) so that cells form a half-confluent monolayer.

After 24 h incubation, aspirate culture medium from the cells. Dispense 100µL of the test solution, including the sample at the appropriate concentration, negative control, positive control, and blank control, into each well. Four different test sample concentrations (100%, 50%, 25%, 12.5%) were tested.

Incubate cells for 24 h (5% CO₂, 37° C, > 90% humidity).

After 24h of testing, the plate and cell morphology were examined under an inverted biomicroscope, and the changes in cell morphology due to cytotoxicity of the sample extract was recorded.

After the examination of the plates, carefully remove the culture medium from the plates. 50 μ l of the MTT solution is then added to each test well and the plates are further incubated for 2 h in the incubator at 37°C. Then the MTT solution is discarded and 100 μ l of isopropanol are added in each well. Sway this plate and subsequently transfer it to a microplate reader equipped with a 570nm filter to read the absorbance (reference wavelength 650nm).

6.4 Data analysis

Compared with blank group, cell survival rate was calculated by following formula.



Viab. (%) =
$$\frac{100 \times OD_{570e}}{OD_{570b}}$$

where: OD_{570e} —is the mean value of the measured optical density of the 100 % extracts of the test sample.

 $\mathsf{OD}_{\mathsf{570b}}$ —is the mean value of the measured optical density of the blanks.

6.5 Microscope evaluation

According to standard, A useful way to grade test samples is given in Table 6-2.

Table 6-2 Qualitative morphological grading of cytotoxicity of extracts

Grade	Reactivity	Conditions of all cultures
0	None	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth
1	Slight	Not more than 20% of the cells are round, loosely attached and without intracytoplasmatic granules, or show changes in morphology; occasional lysed cells are present; only slight growth inhibition observable.
2	Mild	Not more than 50% of the cells are round, devoid of intracytoplasmatic granules, no extensive cell lysis; not more than 50% growth inhibition observable.
3	Moderate	Not more than 70% of the cell layers contain rounded cells or are lysed; cell layers not completly destroyed, but more than 50% growth inhibition observable.
4	Severe	Nearly complete or complete destruction of the cell layers.

6.6 Quality check

A test meets the acceptance criteria if the mean OD_{570} of blanks is ≥ 0.2 .

A test meets acceptance criteria if the 96-well plate left side (row 2) and the right side (row 11) mean of the blanks do not differ by more than 15 % from the mean of all blanks.

6.7 Evaluation criteria

The lower the Viab. % value, the higher the cytotoxic potential of the test item is.

If viability is reduced to < 70% of the blank, it has a cytotoxic potential. The 50% extract of the test sample should have at least the same or a higher viability than the 100% extract; otherwise the test should be repeated.



7 Test result

The qualitative morphological classification of cytotoxicity of extracts from different groups was shown in Table 7-1. The result of cell viability (%) for the test sample extracts at concentrations of 100%, 50%, 25%, and 12.5% was shown in Table 7-2

Table 7-1 Qualitative morphological classification of cytotoxicity of extracts from different groups

Group	Cell morphology observation		
Blank control	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth		
Positive control	Nearly complete or complete destruction of the cell layers.	4	
Negative control	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth	0	
Sample solution (100%)	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth	0	
Sample solution (50%)	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth	0	
Sample solution (25%)	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth	0	
Sample solution (12.5%)	Discrete intracytoplasmatic granules, no cell lysis, no reduction of cell growth	0	

Table 7-2 Optical Density and Viability

Group	Sample solution (100%)	Sample solution (50%)	Sample solution (25%)	Sample solution (12.5%)	Negative control	Positive control	Blank control
Average value	0.4553	0.4558	0.4745	0.4397	0.4642	0.1288	0.4289
SD	0.0351	0.0310	0.0139	0.0282	0.0462	0.0100	0.0395
Survival rate %	106.16	106.28	110.63	102.51	108.22	30.04	100.00

8 Deviations

The test was carried out in strict accordance with the standard operating procedures, and no deviation affecting the validity of the test data occurred.

9 Record Preservation

All raw data and records related to this test and copies of the final report are kept in the archives.



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Test report photo page

Photos and descriptions



Test component description

Random sampling

Model, specification or other description

1

***** End of report *****

